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I, KIM MARSHALL, MANAGER EXAMINATION SUPPORT AND SALES,
hereby certify that the annexed is a true copy of the Provisional specification in
connection with Application No. PP 2902 for a patent by GRADIPORE LIMITED
filed on 9 April 1998.



WITNESS my hand this Twenty-ninth
day of April 1999

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MANAGER EXAMINATION SUPPORT AND
SALES

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AUSTRALIA

Patents Act 1990

GRADIPORE LIMITED

PROVISIONAL SPECIFICATION

Invention Title:

Improved electrophoresis gel and gel-forming apparatus

The invention is described in the following statement:

Technical Field

The present invention relates to electrophoresis gel formation and apparatus suitable for forming gels.

Background Art

5 The preparation of polyacrylamide-based matrices for electrophoresis has conventionally involved the aqueous copolymerisation of acrylamide with a crosslinking agent by free radical chemistry. The free radical polymerisation can be initiated by various processes, and once commenced, the polymerisation reaction proceeds until a gel is formed. Gels are often
10 prepared on an individual basis prior to use, and there can be variations between gels that have been cast separately such that comparison between separations using the same gel type are not reliable. Additionally, there has now been a move to the commercial preparation of preformed gels which should have consistent quality and stable physical characteristics between
15 batches.

Traditionally, polyacrylamide gels have been prepared in glass supports. For commercialisation purposes, synthetic electrophoresis gel supports offer a number of advantages over the traditional supports. These include versatility in processing, light weight properties, improved visual
20 appearance, and shatter resistance.

It has long been recognised that the polymerisation and copolymerisation of acrylamide by free radical chemistry is subject to inhibition by a range of compounds. Specifically, oxygen acts to terminate growing polymer chains resulting in longer polymerisation times. Work by
25 other commercial corporations (Daiichi Pure Chemicals and Novel Experimental Technologies) has recognised the impact of such inhibitors and have attempted to address the issues associated with them.

US patent 5350552 (Daiichi Pure Chemicals) describe a batch process in which polyacrylamide gels for electrophoresis are prepared in a container with a low oxygen atmosphere. The batch process involves placing gel supporting plates into the container, in which they are separated by the aid of partition members. The purpose of the partition members is to act as "polymerisation prevention plates" to assist in enhancing and speeding up the cleaning and processing of the gels with a minimum of gel rejection. The
30 partition members are also used to dissipate the polymerisation exotherm, which is thought to ultimately limit the batch size (up to a maximum of 50
35

cassettes in the batch). The partition members may be made from a variety of synthetic materials such as polyolefins, polystyrene or fluorinated resins, or from rubber, but should be able to "embrace a large amount of oxygen around its surface with high radical absorptivity." The gel support plates themselves may be made out of glass or plastic.

In US patent 5350552, the inventors recognised the requirement of preparing the gels in a low oxygen environment in order to eliminate the appearance of "flaws or stripes" in the gel. The minimisation of flaws in the gel has traditionally been achieved by the use of an overlay solution, which is employed to prevent the re-absorption of oxygen from the atmosphere by the top portion of the gel. The use of such conditions emphasise that oxygen in the container is a problem, and removal of the oxygen is possible either through application of a vacuum or by displacement with an oxygen-free gas such as nitrogen. After the container is filled with nitrogen, the gel solution is introduced. However, it is not specified whether nitrogen flow is maintained during the polymerisation.

US patent 5685967 (Novel Experimental Technologies) describes a process by which a mould for an electrophoresis gel is coated with barrier films, such as silicone oxide, in order to form a polyacrylamide gel suitable for biological separations. Examples are given in the body of this patent in which various plastic materials, coated and uncoated, were examined for their influence on the gel polymerisation, the resultant physical properties and separation. As an illustration, uncoated SAN (styrene-acrylonitrile) cassettes induced poor polymerisation, and correspondingly, poor electrophoresis results. When the SAN material was coated with PET-SiO_x film, the gel quality and performance improved significantly. It was also noted that the oxygen permeability and transmission of the surface in contact with the polymerising solution was an important factor for consideration. This observation is related to an earlier patent specification (WO 90/13020) in which the oxygen permeability of various plastics (PMMA, PET, polystyrene, polycarbonate and polyethylene) and the implication of oxygen in the plastic is discussed.

The present inventors have now obtained improved electrophoresis gels without the aid of barrier films or chemical treatments. Furthermore, large batch production of gels has been achieved by the use of a new gel-forming apparatus.

Disclosure of Invention

In a first aspect, the present invention consists in an apparatus for forming electrophoresis gels, the apparatus including a container having a base and sides, the container being adapted to receive a plurality of gel cassettes; an inlet port positioned in the base of the container and in fluid communication with the chamber; and a baffle positioned over the inlet port, such that, in use, when fluid passes through the inlet port into the chamber, the baffle substantially reduces fluid turbulence and vertical fluid movement in the vicinity of the inlet port during flow of the fluid into the chamber.

The apparatus may be of any configuration, however, the present inventors have found that a container with a substantially square shaped base is particularly suitable. The inlet port is preferably positioned in the middle of the base of the container with the baffle placed directly over the port. Preferably, the baffle has substantially the same cross-sectional shape as that of the container but of smaller dimension to allow fluid to pass around and over the baffle. The baffle is preferably flat and relatively thin in cross-section to minimise flow turbulence as fluid passes around and over the baffle. The baffle is preferably positioned above the inlet port substantially in the same plane, preferably horizontal, as the base.

Fluid may be moved into the apparatus through the inlet port by any suitable means including pumping or gravity feeding.

In one preferred form, the apparatus is placed in a vacuum chamber to assist in the formation of improved gels according to the present invention.

In order to cast a large number of gels in the apparatus, suitable racks which are adapted to hold the cassettes in the correct orientation can be placed in the apparatus.

The apparatus may further include means to control the temperature of the container to assist in the formation of suitable gels. Alternatively, the apparatus may be located in a controlled atmosphere environment.

In a second aspect, the present invention consists in an electrophoresis gel formed by the apparatus according to the first aspect of the present invention.

In a third aspect, the present invention consists in a process of forming an electrophoresis gel in a plastic cassette, the process including the steps of:

(a) pretreating the plastic cassette to substantially remove polymerisation initiators present therein;

- (b) preparing a monomer solution of acrylamides and treating the monomer solution to substantially remove any oxygen or other gaseous polymerisation inhibitors therefrom;
- (c) preparing initiator and co-initiator solutions required to induce polymerisation of the monomer solution, the solutions being treated so as to substantially remove any oxygen or other gaseous polymerisation inhibitors therefrom;
- 5 (d) mixing the monomer solution with the initiator and co-initiator solutions to form an initiated monomer solution;
- 10 (e) applying the initiated monomer solution to the plastic cassette; and
- (f) allowing the initiated monomer solution to polymerise in the plastic cassette.

The cassettes may be manufactured from any suitable synthetic (plastic) material, such as polyesters (PEN, PET, PETG), polyolefins (polyethylene, polypropylene), polystyrene, and any copolymers (SAN). The different materials, however, may require different levels of pretreatment prior to gel formation.

In a preferred embodiment of the third aspect of the present invention, the pretreatment of the plastic cassette is by exhaustive vacuum treatment, 20 optionally with inert gas purging. This can be achieved conveniently using a vacuum chamber in which one or more plastic cassettes are placed. A vacuum is then applied to the chamber with optional inert gas purging, preferably with nitrogen, if required. The time required to substantially remove polymerisation inhibitors will depend on the type of plastic used. 25 The present inventors have found that pretreatment times from 1 to 12 hours have been particularly successful. It will be appreciated, however, that pretreatment times may vary depending on the type of plastic used and the number of cassettes being pretreated.

In order to remove oxygen and other gaseous polymerisation inhibitors 30 from the various solutions, degassing and optional gas purging have also been found to be particularly suitable. In step (c), one means of ensuring the removal of any oxygen or other gaseous polymerisation inhibitors is to treat water, in which the solutions are made, by degassing and optional gas purging prior to adding the initiator and co-initiators to form the solutions.

In a further preferred form, step (e) applying the initiated monomer solution to the plastic cassette is carried out in the apparatus according to the first aspect of the present invention.

5 The gels formed may be continuous or gradient gels comprising standard gel forming ingredients having concentrations of monomer and cross-linker as presently used in standard gels known to the art.

In a fourth aspect, the present invention consists in an electrophoresis gel formed by the process according to the third aspect of the present invention.

10 Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element, integer or step or group of elements, integers or steps but not the exclusion of any other element, integer or step or group of elements, integers or steps.

15 In order that the present invention may be more clearly understood, preferred forms will be described in the following examples with reference to the accompanying drawings.

Brief Description of Drawings

20 Figure 1 is a schematic view of a gel forming tower according to the present invention.

Figure 2 is a diagram showing the positioning of four sets of gel cassettes positioned in a gel forming tower.

25 Figure 3 shows various gels from positions in a gel forming tower shown in Figure 2 Coomassie blue stained after electrophoresis of protein standards.

Figure 4 shows an isotherm (temperature vs time plot) during acrylamide gel polymerisation of a gel positioned at the centre of the gel forming tower during polymerisation.

30 Figure 5 shows an isotherm (temperature vs time plot) during acrylamide gel polymerisation of a gel positioned at the edge of the gel forming tower during polymerisation.

Figure 6 shows the stability of various gels cast in the one batch and stored over a 6 month period, Coomassie blue stained after electrophoresis of protein standards.

Figure 7 shows a comparison of two commercial gels with a gel formed by the present invention Coomassie blue stained after electrophoresis of protein standards.

5 Figure 8 shows a gel stained using silver diamine after electrophoresis of protein standards.

Modes for Carrying Out the Invention

The present inventors have developed a process in which the inhibiting properties of synthetic electrophoresis gel supports or cassettes, such as polyesters (PEN, PETG and PET) and polystyrene and its copolymers (SAN) can be removed by an exhaustive degassing process. Under going 10 such treatment, the gels according to the present invention prepared in plastic cassettes were equivalent to, or in some instances better than the current commercially available gels.

15 Furthermore, as a result of this pretreatment, the present inventors have unexpectedly found the polymerisation process required reduced quantities of initiator and co-initiator, and unexpectedly, the polymerisation exotherm of a gel is much more uniform and controlled. The gels so formed in this controlled exotherm system have greatly enhanced separating properties compared to previous gels made in plastic or in glass cassettes.

20 The process for the manufacture of improved polyacrylamide gels consists of several components. The components and their role in the process is outlined below:

Pretreatment Unit (Vacuum chamber set-up)

This consisted of a high-vacuum chamber, into which high vacuum 25 and an inert oxygen-free gas, preferably nitrogen, was introduced. The gel-forming container which holds the gel cassettes was placed in the chamber and a cycle of vacuum degassing and nitrogen gas purging began (evacuation/nitrogen purge). The pretreatment or degassing cycle may be a single, continuous evacuation, or may be a series of evacuation-purge cycles 30 for a pre-determined length of time.

The pretreatment removes the inhibitors from the plastic cassettes in order to render them suitable for the polymerisation of acrylamide, and different pretreatment times are often required for different plastic cassettes. As an example, cassettes made from the polyesters polyethylene naphthoic acid (PEN) or polyethylene terephthalate-co-glycol (PETG) require degassing 35 times of 1 hour and 2 hours respectively, to produce gels of equivalent

quality. Cassettes prepared from styrene-acrylonitrile copolymer (SAN) require about 12 hours degassing.

During the polymerisation of acrylamide, it is desirable to maintain an inert atmosphere in order to minimise the incorporation of oxygen and other polymerisation inhibitors into the polymer chain. For example, covalently bonded oxygen becomes a weak link in the polymer backbone as it forms a peroxide bond. Without the use of barrier films or chemical scavengers, polymerisation in synthetic gel supports was not previously thought possible, even within a low oxygen environment.

Additionally, gel polymerisations carried out previously required an organic or aqueous overlay, which can interfere with the polymerisation process. The present invention does not require such measures to obtain suitable gels.

Gel Forming Container (Tower)

Previous tower designs have been based on cubic-shaped containers or inverted pyramid designs widening out into cubic shaped area.

A tower for multiple gel casting should satisfy various requirements including having:

minimum hold-up volume
minimum time from the point of initiation to introduction of the solution into the cassettes

non turbulent flow in the tower and cassettes

This has been achieved in the present invention by the use of a tower design which is cubic shaped in which there is a minimum hold-up volume in the area under the level of the plastic cassettes (Figure 1). An inner frame (not shown) on which the cassettes rest may also form part of the tower design, but may not be necessary and its use is dependant upon need and scale. The tower (10) and inner frame may be formed from any material which is not free radical inhibiting and does not interfere with the polymerisation process or solution flow. The tower (10) is preferably made from perspex for ease of processing and visual appeal. It will be appreciated that the tower may also be made of metal including suitable alloys, with or without suitable coatings.

The tower design also encompasses the use of a distributor plate or baffle (12), preferably substantially square in shape with a base (13) and walls (14), of specific dimensions 1/4 to 1/2 way to the cross sectional

diameter of the base of the tower, preferably 1/3 of the way, which is placed over the solution inlet port (12). The use of the plate (12) enables a smooth and even flow of the solutions into the tower by dramatically decreasing the vertical velocity of incoming solutions. Thus, particularly for the formation 5 of gradient gels, disturbance to the pre-formed gradient is minimised. This has been evidenced by dye flow tests in towers prepared in accordance with the present invention. The plate (11) may be fixed to the tower (10) by any appropriate manner, for example, by screws to the base (13) of the tower (10), or to a support bar above, or in any other manner which does not impede the 10 solution flow. The height of the plate (11) from the base (13) of the tower is preferably 3 to 10 mm, and more preferably 5 mm. The plate (11) may be made of a material which does not interact with the solutions or interfere with the polymerisation reaction, and examples of such materials are poly(methyl methacrylate), aluminium, and stainless steel. The plate (11) 15 should preferably be of a minimum thickness, consistent with strength requirements.

Synthetic Electrophoresis Gel Supports (Plastic Cassettes)

The cassettes may be manufactured from any suitable synthetic material, such as polyesters (PEN, PET, PETG), polyolefins (polyethylene, 20 polypropylene), polystyrene, and any copolymers (SAN). The different materials, however, may require different levels of pretreatment prior to gel formation. Most plastics may be used, even those previously highlighted as polymerisation inhibiting plastics and unsuitable like polystyrene. In addition, some materials may require further treatment before use. For 25 example, the polyester PEN may require, but not necessarily require, a mild, alcoholic caustic etch prior to use. It will be appreciated that any other pretreatment step which renders the cassette material more useful in the formation of electrophoresis gels would be included in the scope of the present invention.

Scale of Batch Casting

The batch scale may be readily increased without loss of gel quality. Previously, it has been noted that the batch size for the commercial manufacture of polyacrylamide gels is limited by the resultant 30 polymerisation exotherm. The present inventors have been able to improve on the previous maximum batch size of 50, and were able to produce electrophoresis gels routinely in a batch size of 80. In addition, an increase

in scale by 4, with a batch of 320 gels has also been achieved. It is surprising that the exotherm under this large scale casting was controllable and relatively uniform across the batch.

- The control over the exotherm was evidenced by temperature sensors
5 present within the gel during manufacture at a central and edge position
within the batch.

Correspondingly, the gel quality did not vary across the batch and remained consistent due to the ability to control the exotherm. Experiments performed with other plastics gave similar results for exotherm and other
10 properties, and it appears that batch scale need only be limited by physical practicalities.

Process of Manufacture

Initiator Solutions

During the process, there is preferably a sequential addition of the
15 solution components which comprise the monomer mixture. Free radical polymerisation may be initiated by various processes, but from a commercial point of view, the use of a redox system comprising an initiator and a co-initiator is preferred. In contrast to UV photoinitiation or thermal initiation, a redox system is readily adaptable to multiple gel preparations within a
20 batch process. A common redox system is composed of a peroxide based initiator such as ammonium persulphate or potassium persulphate, and a co-initiating agent, which in conjunction with the initiator, is capable of producing free radicals. Examples of co-initiators are N, N, N', N'-
tetramethylethylenediamine (TEMED) and 3-dimethylaminopropionitrile
25 (DMAPN).

The process involves the addition of the initiators in a manner such that the maximum holding time is achieved, allowing the solutions to flow into the container and cassettes before the onset of polymerisation. This has the effect of causing:

- 30 minimum disturbance of solution flow
minimum disturbance of the desired monomer gradient for gradient gels
minimum batch rejection due to premature polymerisation
The preferred ratio between the initiator and co-initiator components
35 is 1:1. It will be appreciated, however, that other ratios such as 2:1 and 1:2 can be used.

Monomer Solutions

The need to degas the monomer solutions prior to use has long been recognised as a necessary step for the formation of clear and reproducible polyacrylamide gels free of defects. The solutions may be degassed by 5 evacuation using a vacuum pump or water aspirator, or the solutions may be bubbled with an inert gas such as argon, helium or nitrogen, until a very low level of dissolved oxygen is reached.

The use of an improved process incorporating pretreatment of the plastic cassettes, an inert atmosphere for electrophoresis gel formation and 10 degassed solutions enable very low initiator levels to be used when forming the gels in plastic cassettes. Concentrations typically used in acrylamide polymerisation are in the range of 1 to 10 mM. While initiator levels of less than 1 mM are achievable using glass cassettes, the same low concentrations have previously been found not able to yield good polymerisation in 15 synthetic materials without the use of barrier films or chemical scavengers. With this improved process according to the present invention, initiator concentrations of less than 1 mM are routinely used.

The use of low initiator levels has enabled the production of 20 polyacrylamide gels in plastic cassettes with improved qualities with respect to:

- control over the polymerisation exotherm
- storage stability
- silver staining
- protein and other biomolecule separation

The use of high levels of initiator within the process has been shown to 25 cause the monomer solution to polymerise rapidly, producing an uncontrollable exotherm and brittle gels from the evaporation of water from the gel. Additionally, the incorporation of initiator derived fragments (usually SO_4^- , if persulphate is used) into the polymer chain introduces 30 charged groups into its structure, which is likely to interfere with the separation of biomolecules, affect the level of sensitivity achieved with silver staining, and influence the matrix stability by catalysing hydrolysis.

Therefore, by minimising the presence of oxygen and other volatile 35 inhibitors, the level of initiator may be adjusted down to very low levels while maintaining satisfactory rates of polymerisation for a commercial scale. While the decrease in oxygen has been recognised to increase the rate of

polymerisation, previous practice is not to decrease initiator levels accordingly, as the importance of initiator end groups in the polymer structure has not been recognised.

METHODS

5 Plastic Cassette Preparation

Using SAN cassettes as an example, the pretreatment consisted of subjecting the SAN cassettes to three evacuation/nitrogen purge cycles over a period of 1 hour. Degassing of the cassettes under high vacuum was then left to proceed overnight (12 hours). The cassettes were then subjected to a further three evacuation/nitrogen purge cycles, and then equilibrated to atmospheric pressure under an atmosphere of nitrogen.

10 Solution Preparation

The required amount of acrylamide and crosslinking agent (generally N, N'-methylene bisacrylamide) to give the desired %T and %C ratio was dissolved in water. To this mixture was added an aliquot of Tris hydrochloride buffer to achieve a final buffer concentration of 0.375M. The solution was then adjusted to pH 8.8 and made up to the final desired volume with water. The monomer solutions were then degassed by bubbling gently with nitrogen gas until less than 1% dissolved oxygen was obtained.

15 Monomer solutions were maintained under an nitrogen atmosphere during gel manufacture.

20 The water used to prepare the individual components of the redox initiator system, ammonium persulphate and TEMED, was degassed in the same manner. When the desired oxygen level was achieved, the initiator solutions were then prepared.

25 Gel Preparation

With the aid of peristaltic pumps, the individual monomer solutions, the initiator and co-initiator solutions were pumped into the tower in a sequential manner. The monomer solutions were mixed in-line first, the co-initiator was then added in-line, and finally, the initiator was introduced in-line, prior to reaching the cassettes within the tower and vacuum chamber. The initiated monomer solution was then pushed up to the required level in the cassettes with the aid of a salt solution. Once in the cassettes, the solution was left to polymerise under an atmosphere of nitrogen, over a period of 2 hours. Once this time had elapsed, the gels were removed from the tower, and placed into a 60°C oven for 1 hour for a post-polymerisation

curing step. Alternatively, the gels may be left in the tower and exposed to elevated temperatures *in situ* for curing if required. Once cured, the gels were placed in an 18°C room, and left to cool to room temperature overnight.

EVALUATION OF THE RESULTANT POLYMER MATRICES

5 Plots were made to show the exotherm is controllable throughout the batch of 320 cassettes (no more than 45°C).

Quality across the batch is maintained

Storage stability trials showed the gels were capable of a shelf-life of at least 6 months at 4°C, while the shelf-life of other commercially available 10 gels in synthetic cassettes was 3 months at 4°C. Similar gels in glass cassettes have been found to have a shelf-life of 30 days at 4°C.

Separation - was at least equivalent or superior to that of other commercial gels.

15 Silver staining - was shown to be equivalent to, or better than that of other commercial gels

The ability to prepare gels in a variety of different synthetic materials without loss of gel performance was demonstrated.

SUMMARY

20 Polyacrylamide matrices suitable for electrophoresis were prepared by the present inventors in synthetic gel supports without the use of barrier films or chemical scavengers.

The improved method for the preparation of the polyacrylamide gels according to the present invention can be readily scaled up for mass production in a batch process.

25 The improved batch process incorporates several features which enabled high quality polyacrylamide gels to be prepared in a reproducible manner using a variety of synthetic supports with minimum batch rejection.

The polyacrylamide gels prepared with the improved process had fewer faults in the polymer structure as there was no or minimal 30 incorporation of oxygen, other inhibitors or initiator-derived fragments into the polymer chain.

As a result, the polyacrylamide gels formed using the improved process had a number of improvements over other gels, with respect to:

- control over the polymerisation exotherm
- 35 • quality
- storage stability

- separation
- silver staining

It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention as shown in
5 the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

Dated this ninth day of April 1998

GRADIPORE LIMITED
Patent Attorneys for the Applicant:

F B RICE & CO

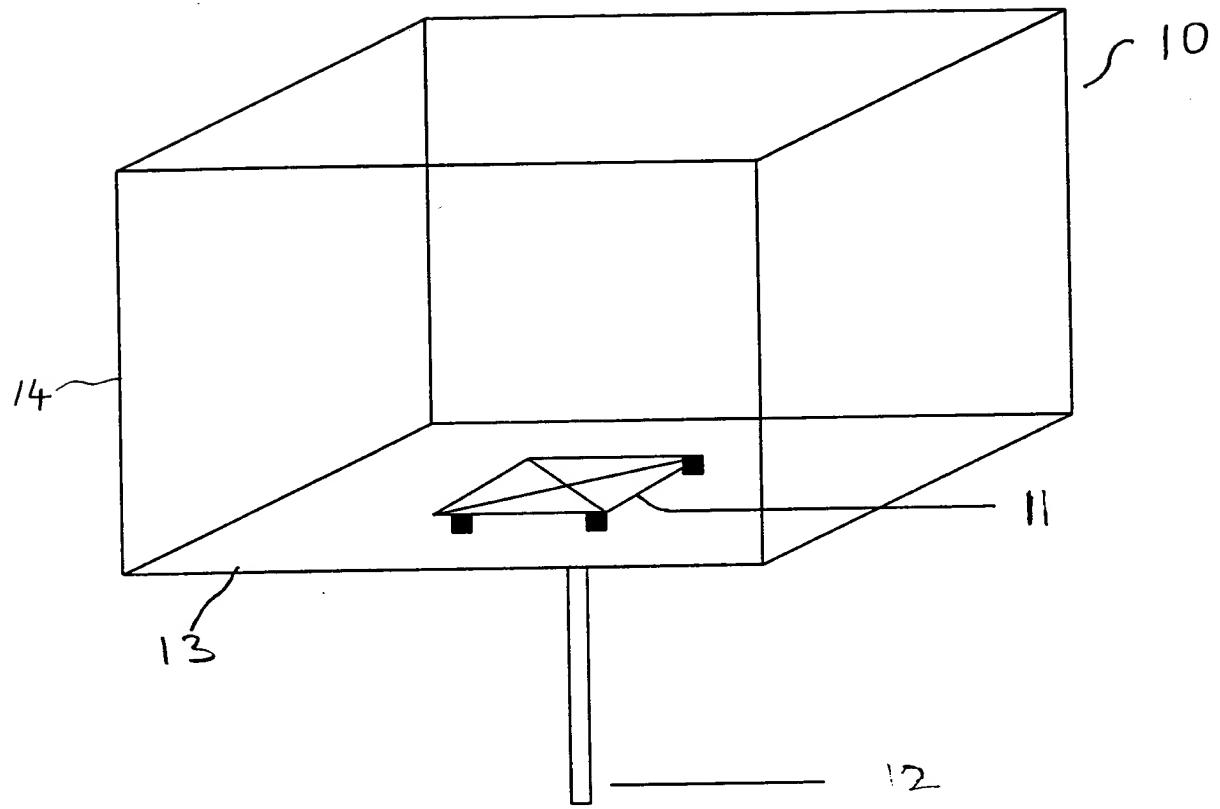


Figure 1

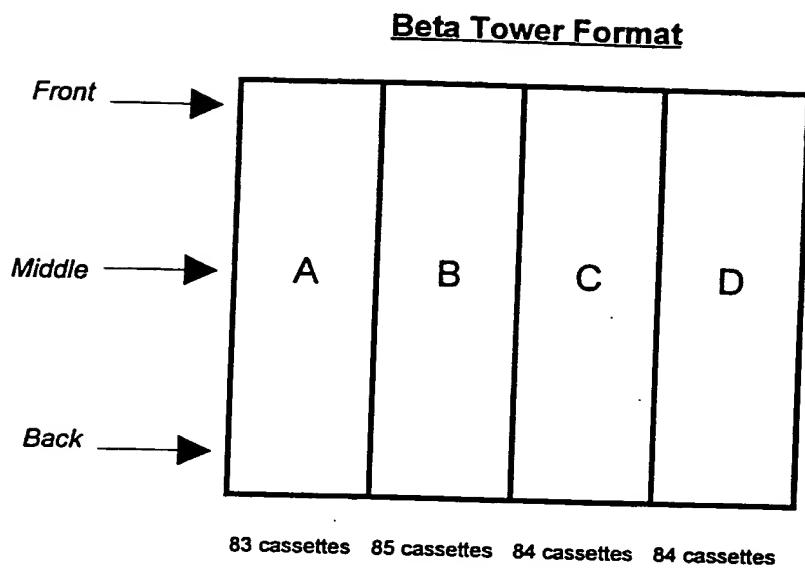


Figure 2

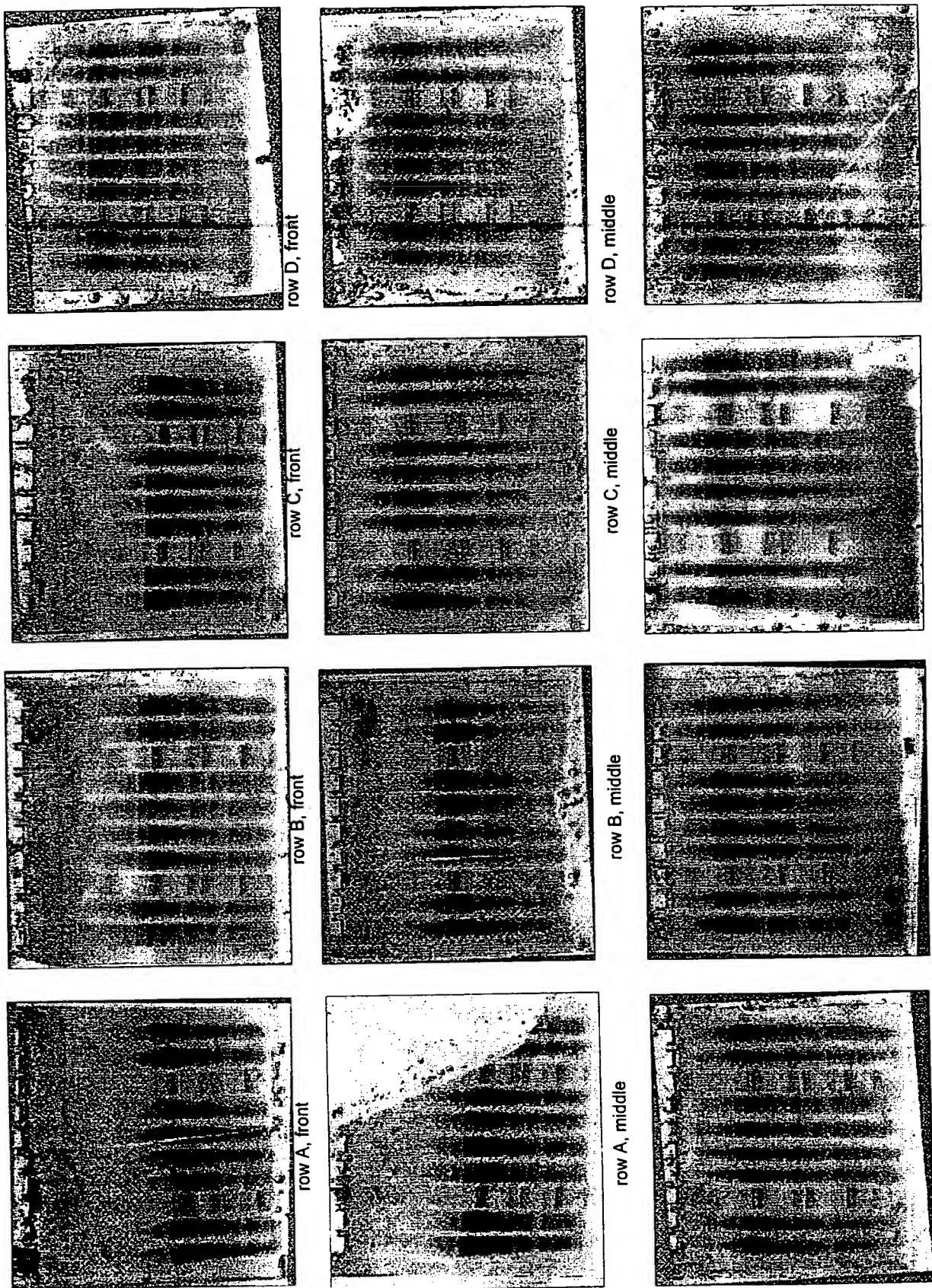


Figure 3

row B, back

row A, back

row D, back

row C, back

Variation in gel temperature during polymerization - centre

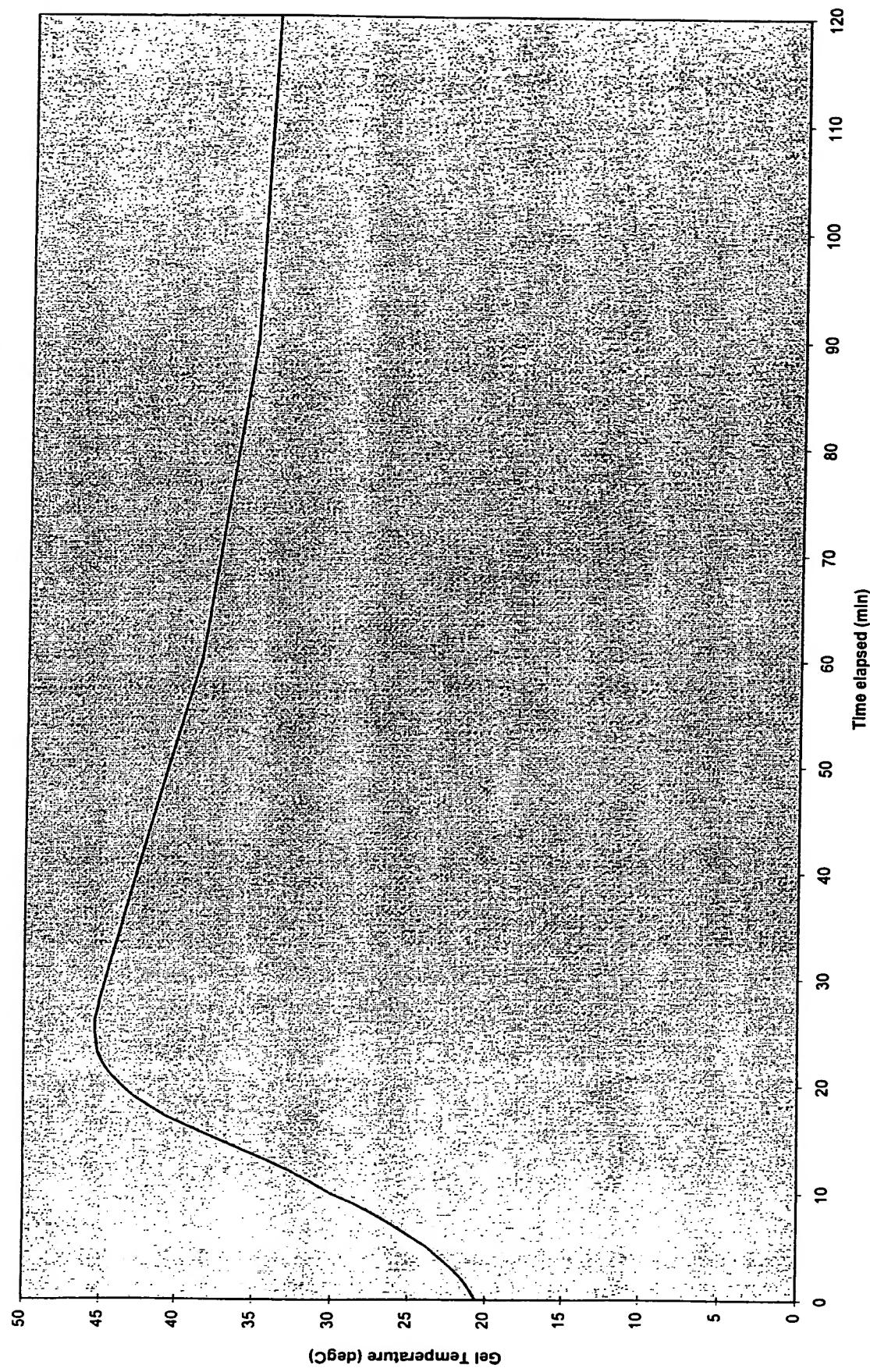


Figure 4

Variation in gel temperature during polymerization - edge

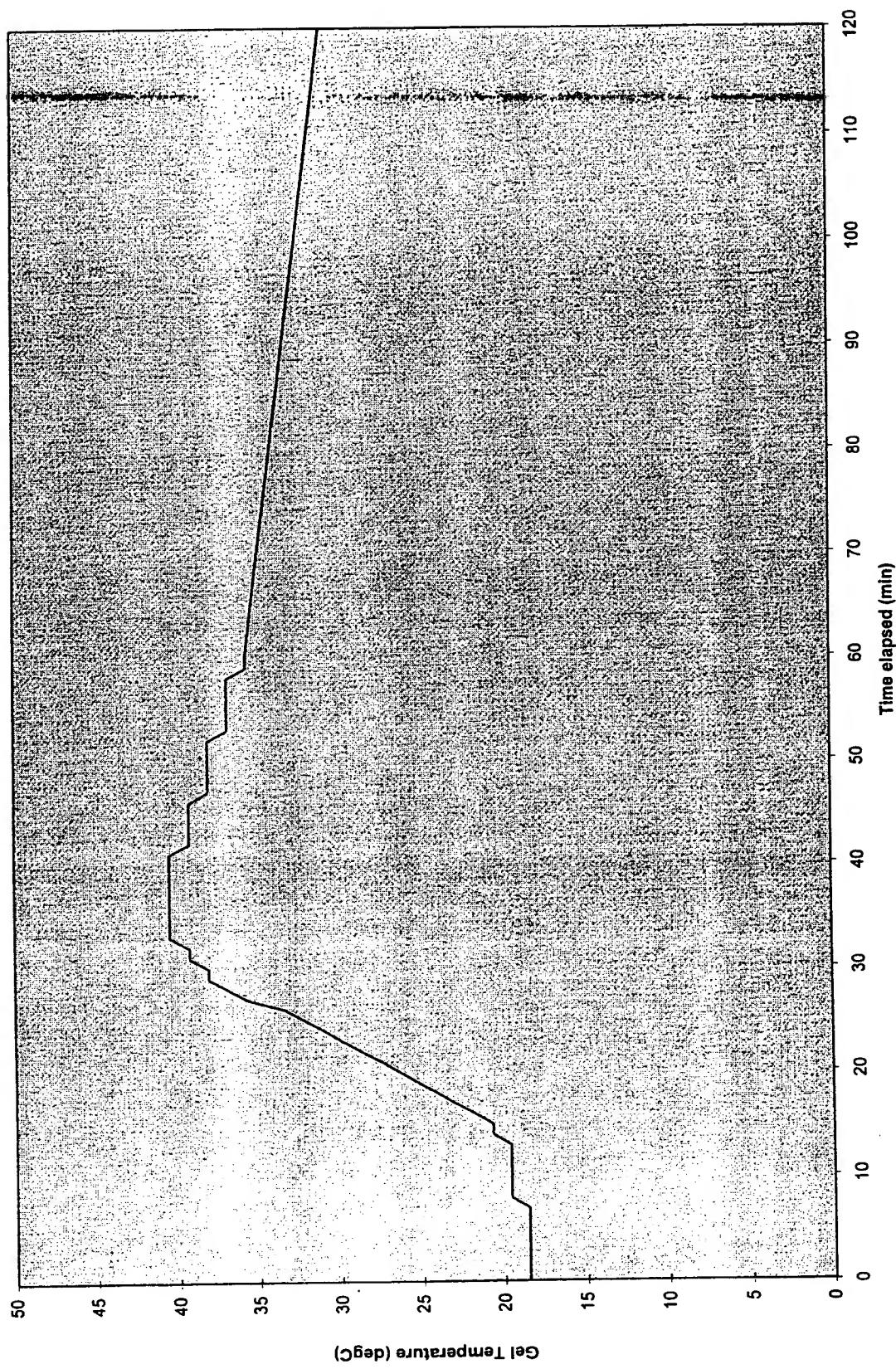
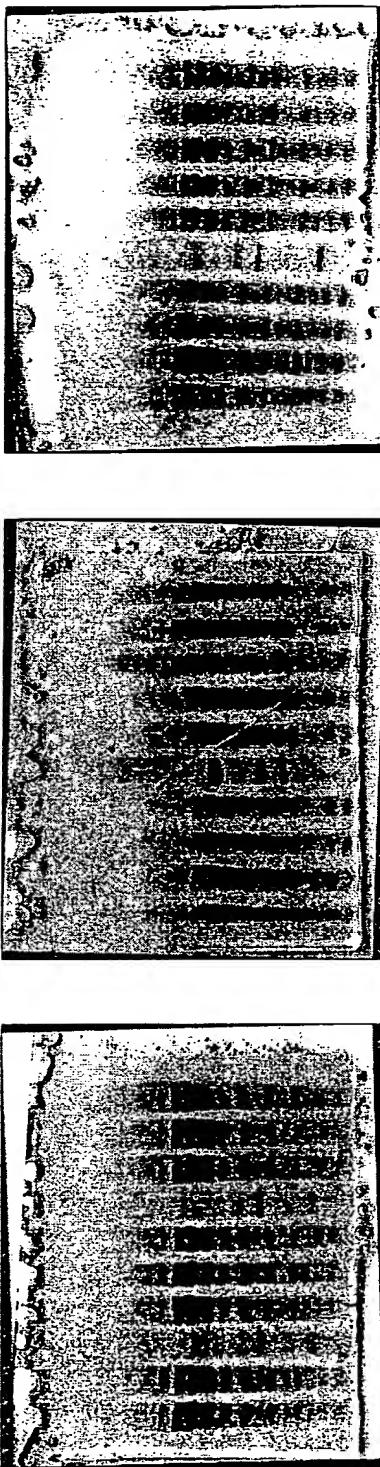


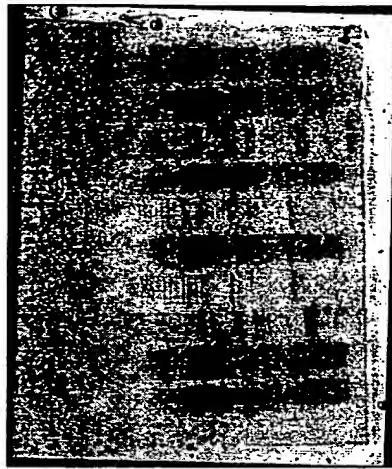
Figure 5

Gel Stability

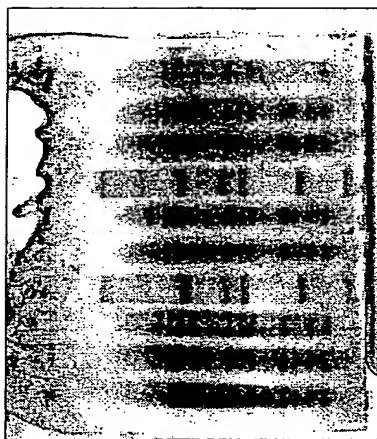


Day 1

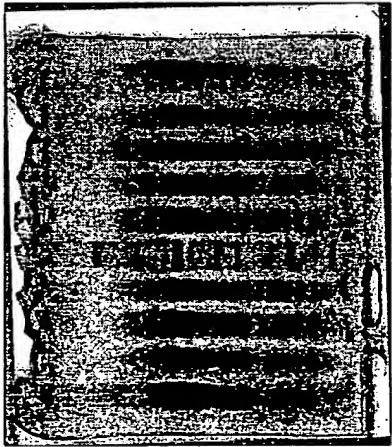
2.5 months



3.5 months



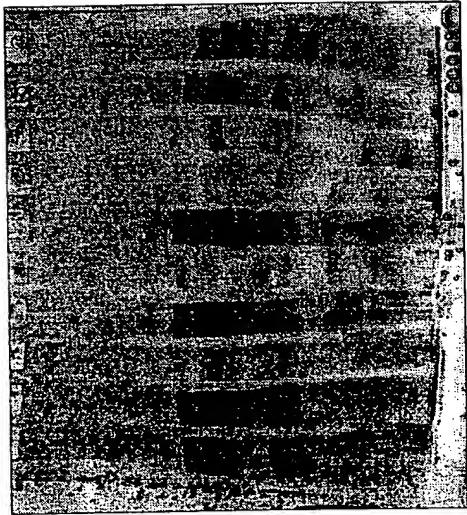
5 months



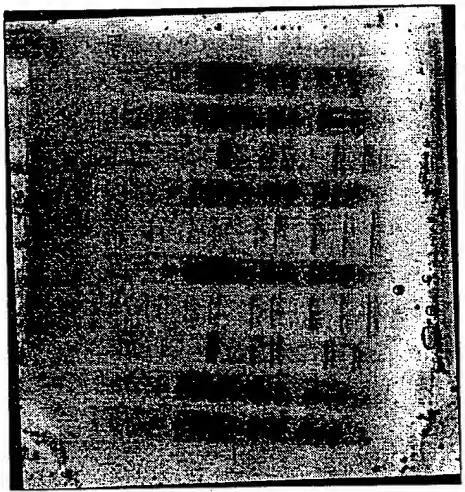
4.5 months

Figure 6

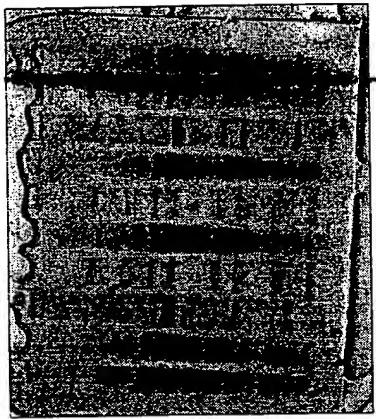
4-20% TG Gel Comparison



Commercial gel 1



Commercial gel 2



Test gel

Figure 7

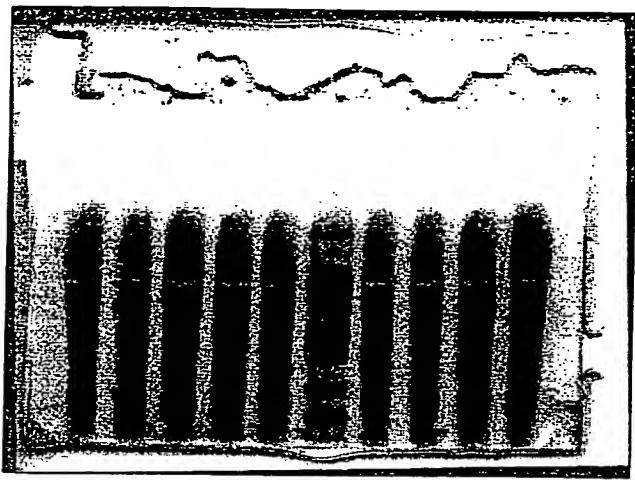


Figure 8

